

# POPULATION ASSAY: PROSPORE AMPOULE

LOT#: \_\_\_\_\_ LABELED POP/POP LEVEL: \_\_\_\_\_

ORGANISM(circle one): *G. stearothermophilus* Other \_\_\_\_\_

Fill Volume (circle one): 1ml 4 ml 1.9 ml SPS Other \_\_\_\_\_ ml

## PROCEDURE:

1.0 Vortex the ampoule for 1 minute in an Upside Down Position, and then for 1 minute in Normal Vertical Position being careful to wash out any spores that may be adhering to the glass in the upper tip of the ampoule. This step is necessary to achieve an accurate population count. Then aseptically transfer a 1ml aliquot to a sterile, screw-capped, 10 ml test tube containing 9 ml of sterile, processed water.

2.0 Heat shock in a water bath (10 minutes at 80°-85°C for mesophiles and 15 minutes at 95° - 100°C for *G. stearothermophilus* and other thermophiles). Immediately cool in a water bath of 0° - 4°C.

Start Time/Temperature: \_\_\_\_\_ / \_\_\_\_\_ °C End Time: \_\_\_\_\_

Initial and Date: \_\_\_\_\_ / \_\_\_\_\_

3.0 Vortex the tube for 15-20 seconds.

4.0 Perform serial dilutions by pipetting out 1.0 ml of the aliquot into another sterile, screw-capped 10 ml test tube containing 9.0 ml of sterile, processed water. Repeat from step 3 until desired dilution factor is reached.

5.0 From the next-to-the-last dilution, pipette out 1.0 ml into each of three Petri plates. Repeat for the final dilution.

6.0 Within 20 minutes, add approximately 20 ml TSA, pre-sterilized and cooled to 47° ± 2°C. Swirl to distribute spores evenly in agar and allow to solidify. Also pour 1 Media Negative Control Plate.

TSA Temperature: \_\_\_\_\_ °C TSA Lot #: \_\_\_\_\_

Initial and Date: \_\_\_\_\_ / \_\_\_\_\_

7.0 Invert and incubate the plates (30°-35°C for mesophiles, 55°-60°C for *G. stearothermophilus* and other thermophiles). Incubate Media Negative Control at same temperature as assay.

Incubation Start Time: \_\_\_\_\_ Incubator #: \_\_\_\_\_

Initial and Date: \_\_\_\_\_ / \_\_\_\_\_

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8.0 Examine all plates at 24 ( ± 1) hours and record the number of colony forming units (CFU's) per plate. Calculate the average number of CFU's per carrier by using the formulas below.

Total @ 24 hrs / number of plates counted x DF x AVF = CFU/ampoule  
 DF= Dilution factor (absolute value of the reciprocal of the dilution)  
 AV= Average number of colonies per ampoule  
 AFV=Ampoule fill volume

**Incubation End Time/Initial & Date:** \_\_\_\_\_/\_\_\_\_\_

### CFU COUNTS AT 24 HOURS

# dilutions \_\_\_\_\_

#### **24hrs**

Plates 1. \_\_\_\_\_ 2. \_\_\_\_\_ 3. \_\_\_\_\_ Total @ 24hours: \_\_\_\_\_

Total @ 24 hrs \_\_\_\_\_ / 3 x \_\_\_\_\_(DF) x \_\_\_\_\_(AFV) = \_\_\_\_\_(AV)CFU/ampoule  
 (4 decimals)

### CFU COUNTS AT 24 HOURS

# dilutions \_\_\_\_\_

#### **24hrs**

Plates 1. \_\_\_\_\_ 2. \_\_\_\_\_ 3. \_\_\_\_\_ Total @ 24 hours: \_\_\_\_\_

Total @ 24 hrs \_\_\_\_\_ / 3 x \_\_\_\_\_(DF) x \_\_\_\_\_(AFV) = \_\_\_\_\_(AV)CFU/ampoule  
 (4 decimals)

# of Dilutions = Dilution Factor

- 1 = 10
- 2 = 100
- 3 = 1000
- 4 = 10000
- 5 = 100000
- 6 = 1000000

TFTC = Too few to count <10CFUs  
 TNTC = Too numerous to count >300CFUs

Media Negative Control: \_\_\_\_\_

Sum of the AV of both dilution / 2 =CFU/ampoule

\_\_\_\_\_ / 2 =

\_\_\_\_\_ x10 \_\_\_\_\_ CFU/ampoule (4 decimals)

Read By: \_\_\_\_\_ Date: \_\_\_\_\_

Reviewed By: \_\_\_\_\_ Date: \_\_\_\_\_